

# **Institute for Ecology of Industrial Areas in Katowice**

Chosen physiological parameters analysis of Miscanthus x giganteus and Spartina pectinata cultivated on soil contaminated with heavy metals. Characterization of measuring equipments

Szymon Rusinowski, MS

Katowice, 20 October 2016

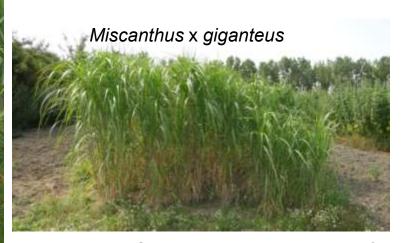
### Presentation Plan

- Presentation of results form preliminary studies
  - Background
  - Aim of studies
  - Site description
  - Experiment design
  - Material & methods
  - Results
  - Conclusions
- Presentation of measuring equipment used in research by IETU Environmental Remediation Team:
  - Infra-Red Gas Analyser (IRGA) "Lc ProSD, ADC Bioscientific, UK"
  - Fluorimeter " Handy Plant Efficiency Analyser, Hansatech Instruments ltd., UK
  - Plant pigments content meter "Dualex Scientific +, Force A, France"
  - Ceptometer (LAI-meter) "LP-80, Decagon Device, USA"



# **Energy Crops**

- Despite the fact that energy crops are not hyperaccumulators, due to its huge biomass production ability they can extract totally a lot of heavy metals
- European Union Directive 2009/28/EC (RES) indirectly prescribe lands categories, where energy crops can be cultivated



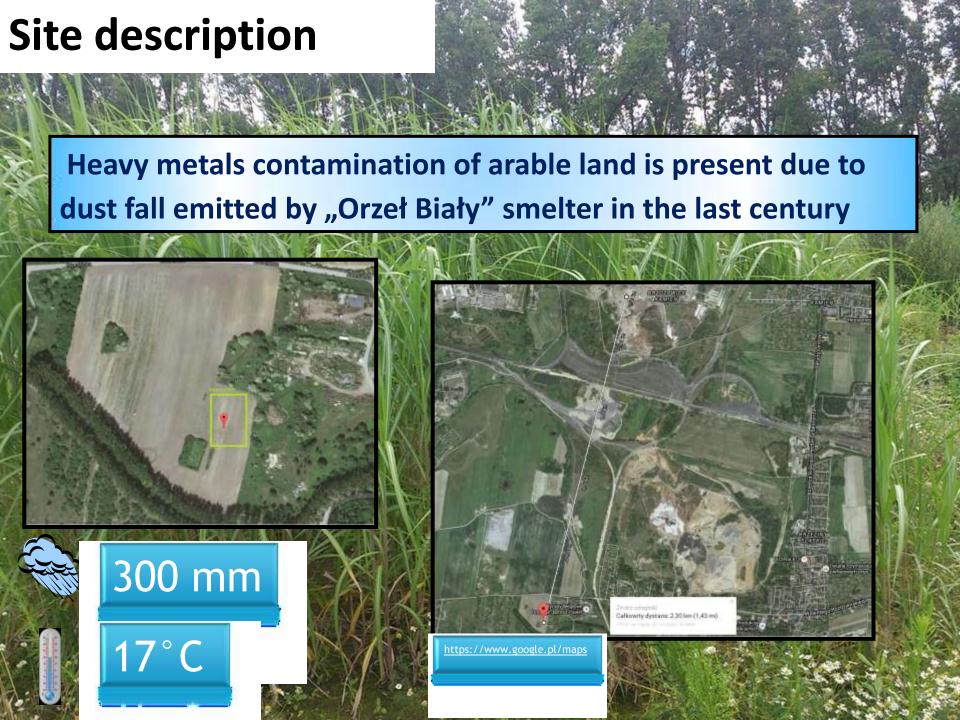


 Miscanthus x giganteus and Spartina pectinata are second generation energy crops

# **Aim of Study**

The aim of preliminary studies was to analyze chosen physiological parameters, mostly associated with photosynthetic aparatus of Spartina pectinata and Miscanthus x giganteus cultivated on soil contaminated with Cd and Pb additionally treated with different fertilizers





# **Experiment design**



- Miscanthus x giganteus (M)
- Spartina pectinata (S)

Three experimental options were set up:

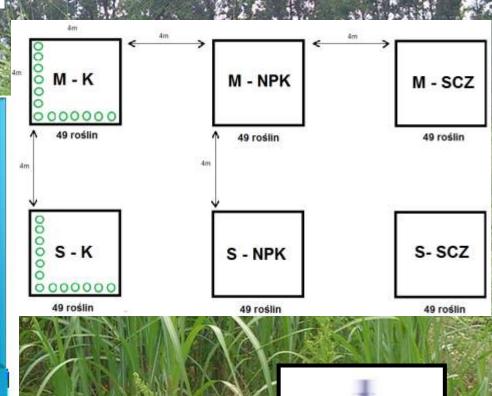
- Control (K)
- Chemical fertilization NPK (NPK)
- Fertilization with commercialy alavailable microbial inoculum (SCZ)

alavailable microbial inoculum (SCZ)

Fertilization with commercialy

Three randomly selected plants not exposed to edge effect from each plot were analyzed. (pseudo-replication)

(pseudo-replication)



http://static.shoplo.com





Soil Pb and Cd content(FAAS)

Gas exchange measurments (LC Pro +, ADC Bioscientific )

Chlorophyll content measurments (CL-01, Hansatech Instruments Ltd.

Chlorophyll a fluorescence measurments (Pocket)
PEA, Hansatech
Instruments Ltd. )

Leaves Pb and Cd content (FAAS)







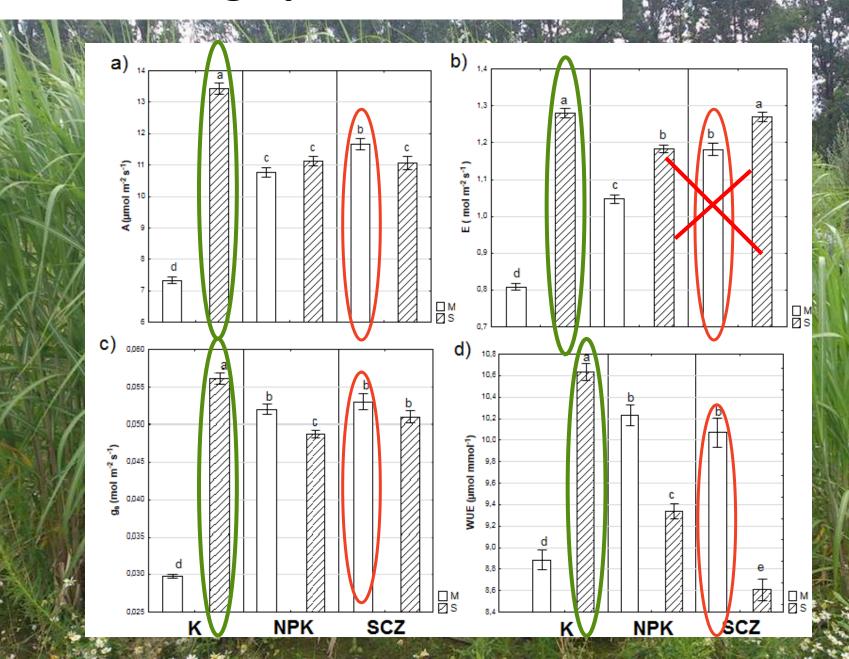
# **Soil characteristic**

| 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1                 |                     |                  | STREET, MANAGEMENT | 3 V2 C            | 超了各 卡罗             | a it lesse        |
|---|---------------------|------------------|--------------------|-------------------|--------------------|-------------------|
|   | EXPERIMENT VARIANTS |                  |                    |                   |                    |                   |
| Parameters  | M-K                 | M-NPK            | M-SCZ              | S-K               | S-NPK              | S-SCZ             |
| Total Cd and Pb content (mg kg-1)                     |                     |                  |                    |                   |                    |                   |
| Cd  | $18,32 \pm 0,99a$   | 19,49 ± 0,65a    | 19,37 ± 0,59a      | $12,64 \pm 0,53b$ | $13,04 \pm 0,42b$  | $12,96 \pm 0,530$ |
| Pb  | $429,89 \pm 4,27a$  | 466,29 ± 12,35a  | 441,51 ± 19,42a    | 282,13 ± 14,69b   | 287,49 ± 8,57b     | 278,45 ± 10,115   |
| Bioavailable Cd and Pb content (mg kg <sup>-1</sup> ) |                     |                  |                    |                   |                    |                   |
| Cd  | $0,72 \pm 0,06$ b   | $1,03 \pm 0,03a$ | $0.76 \pm 0.07$ b  | $1,00 \pm 0,09a$  | $1,00 \pm 0,07a$   | $1,02 \pm 0,02a$  |
| Pb  | $1,17 \pm 0,04b$    | $1,38 \pm 0,05a$ | $1,36 \pm 0,07a$   | $1,38 \pm 0,02a$  | $1,45 \pm 0,05a$   | $1,49 \pm 0,04a$  |
| Soil physico-chemical parameters                      |                     |                  |                    |                   |                    |                   |
| Humidity(%)   | $4,54 \pm 0,56b$    | 6,06 ± 1,05ab    | 6,20 ± 1,32ab      | 8,21 ± 0,61a      | $6,86 \pm 0,63$ ab | 7,11 ± 0,60a      |
| рН  | $6,97 \pm 0,06a$    | $6,72 \pm 0,06b$ | 7,07 ± 0,10a       | 6,66 ± 0,08b      | $6,45 \pm 0,03c$   | $6,40 \pm 0,02c$  |
| EC (μS)   | 79,88 ± 7,12a       | 79,30 ± 5,24a    | 90,20 ± 8,95a      | 75,39 ± 1,34a     | $83,83 \pm 2,05a$  | 79,48 ± 0,41a     |

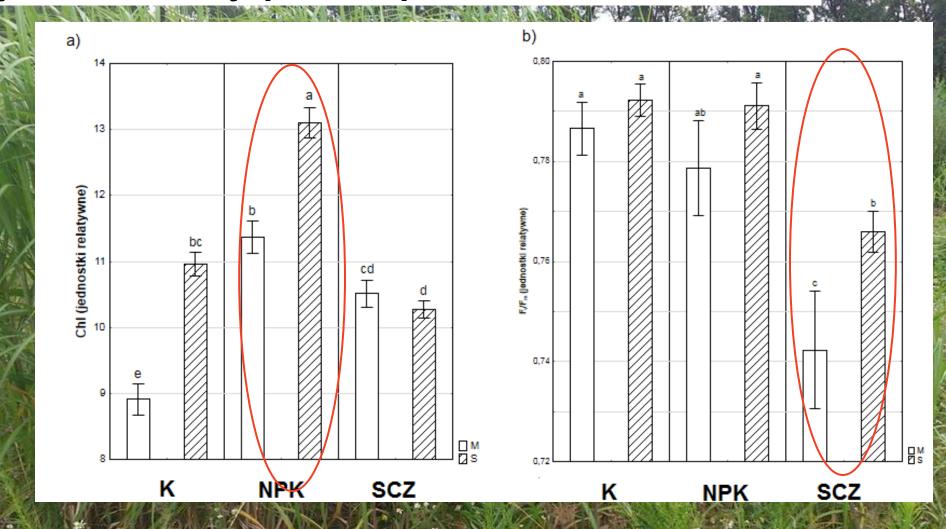
### **Leaves Pb and Cd accumulation**



# Gas exchange parameters



# Leaves chlorophyll content and general plant vitality (Fv/Fm)

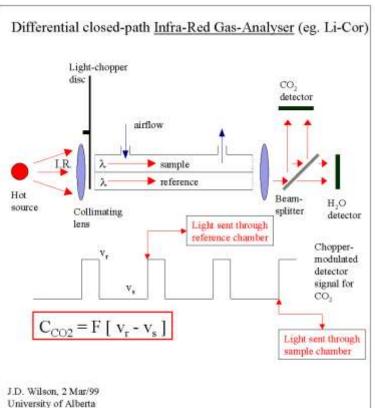


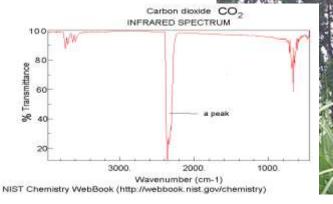
# Conclusion

- \* Influence of fertilizers on plants is species specific, photosynthesis process is more intensive in fertilized *Miscanthus* x *giganteus* when compare to control, however for *Spartina pectinata* opposite phenomenon was observed
- \*Microbial inoculum caused higher Cd leaves accumulation in both plant species
- \* Chemical fertilization significantly increase chlorophyll content in both species
- \* Miscanthus x giganteus accumulates more Cd and Pb in leaves than Spartina pectinata
- \* Despite the Miscanthus accumulate more Pb and Cd in leaves there were no visible effects of those elements toxicity on photosynthetic apparatus

### Infra-Red Gas Analyzer

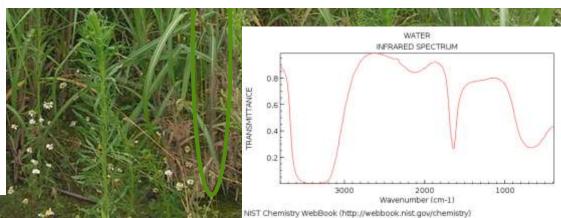
#### What informations are obtained?





### **Measured parameters:**

- Photosynthesis intensity (A)
- Substomatal CO<sub>2</sub> (C<sub>i</sub>)
- Transipration rate (E)
- Stomatal conductance (g<sub>s</sub>)
- Photosynthetic active radiation (PAR)

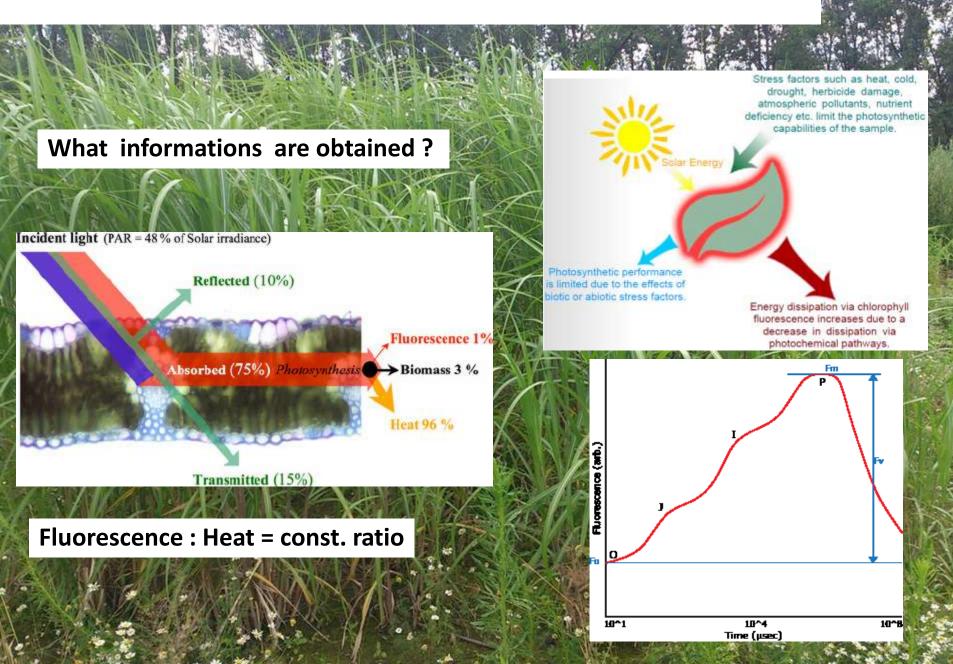


# Infra-Red Gas Analyzer



- Possibility to control: Light intensity, CO<sub>2</sub> concentration, temperature in leaf chamber
- Possibility to measure all parameters on leaves with different size and morphological structure
- Measurment is performed on leaf sample in situ

### Fluorimeter (Chlorophyll a fluorescence)

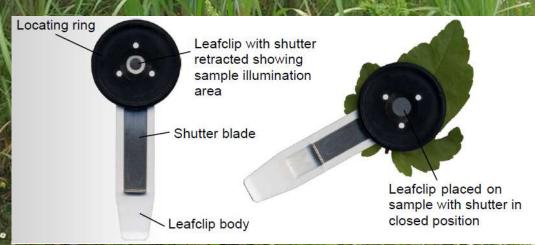


### Fluorimeter

How we measure?



- Samples required dark-adaptation (25 – 40 min)
- 108 measurments in 1 sec
- Derived informations about lightdepending photosynthesis phase



## Chlorophyllmeter

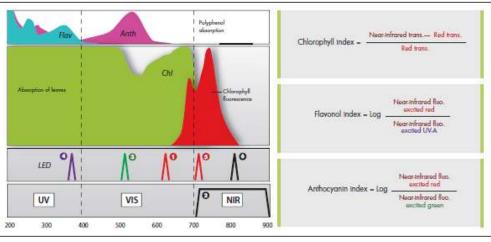
#### What informations are obtained?

#### **POLYPHENOLS** measurement

Near-infrared chlorophyll fluorescence (2) is measured under a first reference excitation light (2) not absorbed by polyphenols. It is compared to a second sampling light specific to a particular type of polyphenols (e.g. green (2) for anthocyanins or UV-A (2) for flavonols). Only a fraction of this light reaches the chlorophyll in the mesophyll and can generate near-infrared fluorescence.

#### CHLOROPHYLL measurement

The leaf chlorophyll content can rapidly and accurately be assessed from light transmission. A first wavelength very close to the red quantifies the chlorophyll and a second in the near-infrared ocn take into account the effects of leaf structure.



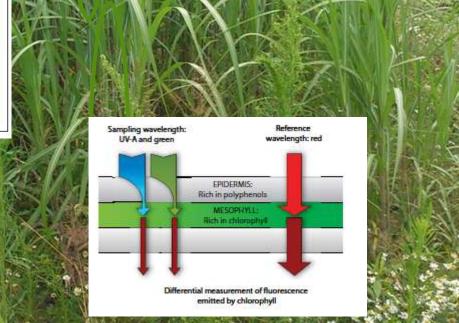
(1) Optimal conditions: Photosynthesis

(2) Nitrogen deficiency:

Secondary metabolism

### Measured parameters:

- Chlorophyll content (Chl)
- Flavonoids content (Flav)
- Anthocyanins content (Anth)
- Nitrogen Balance Index (NBI)



# Chlorophyllmeter

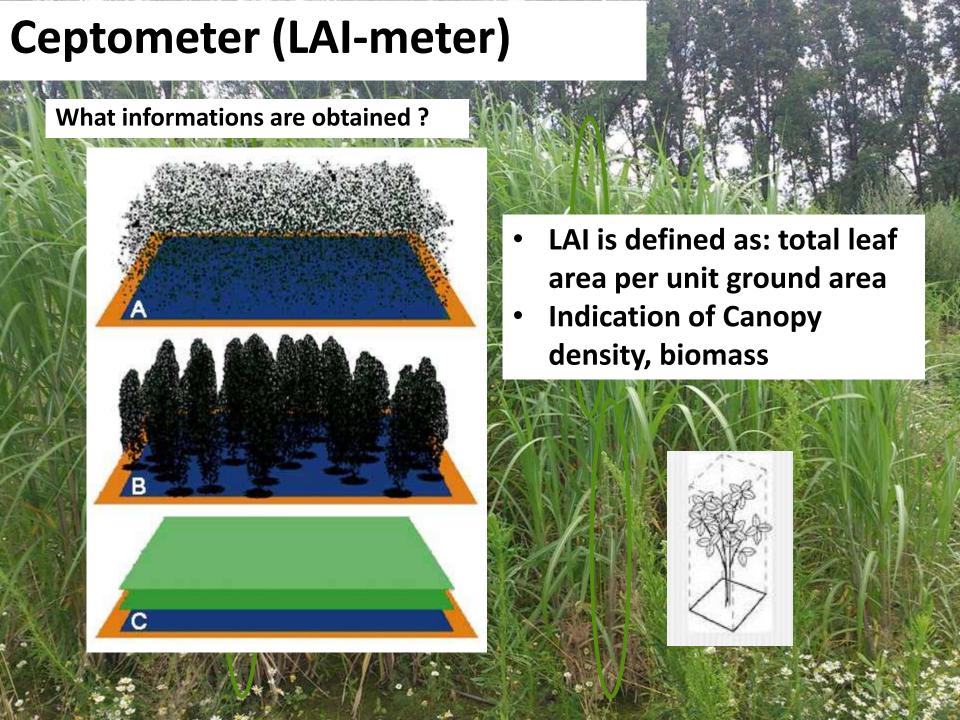
#### How we measure?

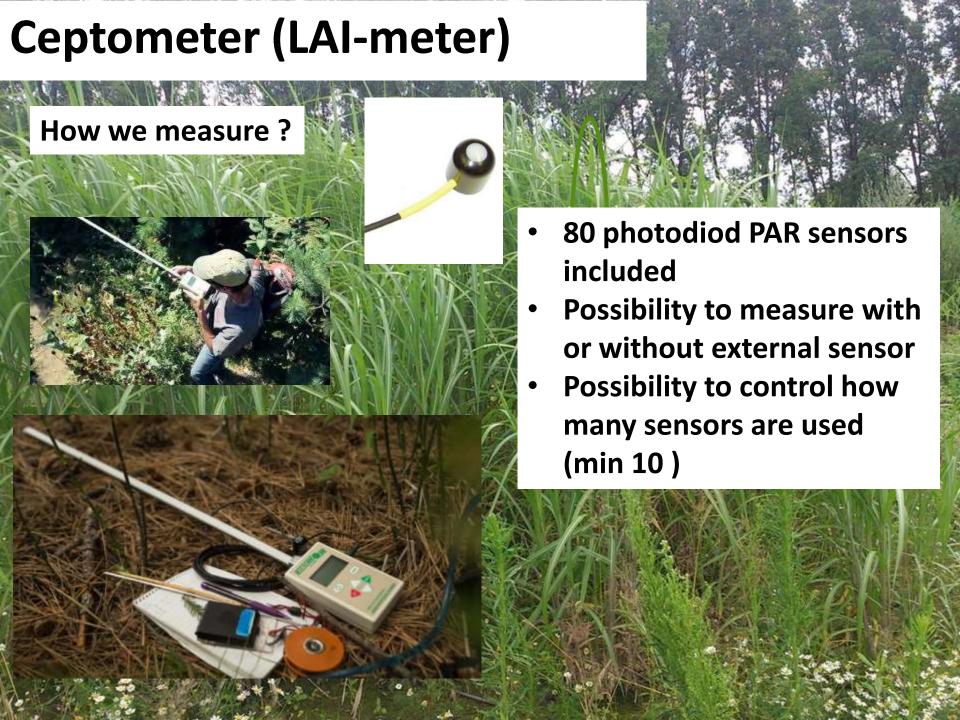


- 1 measurement = 1 sec
- 4 parameters = 1 measurement
- Included GPS (< 2,5 m resolution)</li>
- 10 000 measurements memory

3 main rules to respect to compare treatments :

- Make readings on leaves with the same physiological age. Prefer well developed young leaves.
- Make readings on the same position of the leaves, for all leaves
- The entire Dx window must be covered by the leaf.







## Thank you for your attention

mgr Szymon Rusinowski rusinowski@ietu.katowice.pl